#### Preliminary Experiments to Assess the Oncogenicity of Cellular DNA

Keith Peden, Li Sheng, Achintya Pal, and Andrew Lewis

Division of Viral Products

Office of Vaccines Research and Review

CBER, FDA

Don Blair, Meropi Athanasiou, Stephen Hughes, and John Coffin

NCI - FCRF

Ron Honchal and Frank Sistare CDER, FDA

#### Development of Quantitative Assays to Evaluate Potential Risks Associated with Residual Cell-Substrate DNA

### Projects Underway in OVRR/CBER to Assess the Risks Associated with Residual Cell-Substrate DNA

 Development of in vivo Assays to Assess the Oncogenicity of DNA

Li Sheng, Achinto Pal, Keith Peden, Andrew Lewis (CBER) Ron Honchal and Frank Sistare (CDER) Don Blair, Meropi Athanasiou, Steve Hughes, John Coffin (FCRF-NCI)

 Development of in vitro Assays to Assess the Infectivity of DNA

Li Sheng and Keith Peden (CBER)

## Operational Principles for Regulatory Decisions Regarding Cell-Substrate DNA

- Evaluations of risk need to be based on experimental data on the biological activity of DNA
- As human human immunization data are unattainable, it is prudent to make estimates based on the <u>most</u> sensitive model systems
- As more data are obtained, risk estimates may change

## Development of a Sensitive Animal Model to Assess Oncogenicity of DNA

To develop a quantitative *in vivo* assay to measure the oncogenicity of DNA:

- Choose oncogenes that have been shown to transform efficiently primary cells in culture
- Express these oncogenes under promoters known to function efficiently and for prolonged periods in mice
- Design a "modular" expression plasmid, such that other oncogenes can be evaluated as necessary

## Development of a Sensitive Animal Model to Assess Oncogenicity of DNA

#### Expression Vector

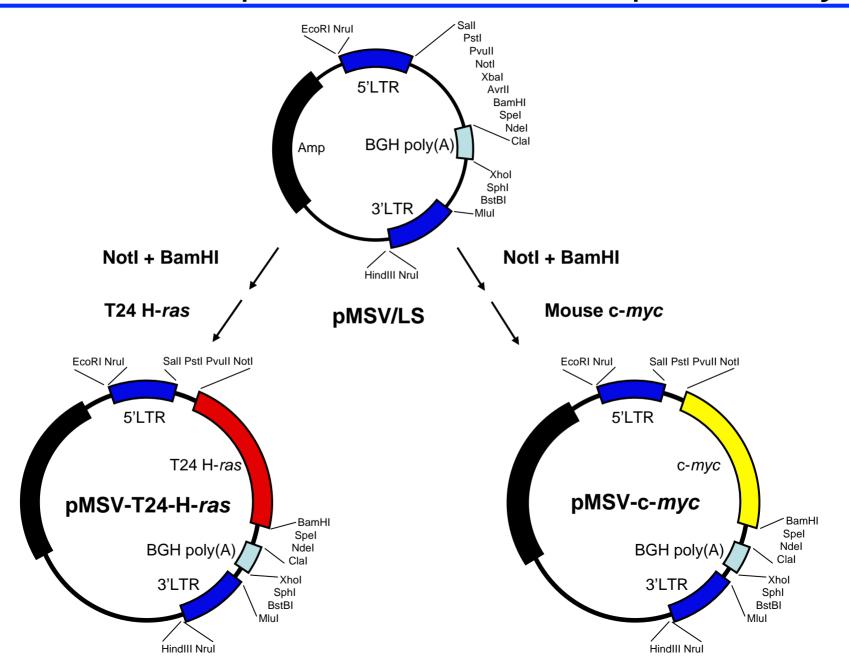
- Promoter is the 5' MSV LTR
- poly(A) site is from BGH
- Second terminator is the 3' MSV LTR
- Backbone pLS3 (pUC19 derivative)

# Development of a Sensitive Animal Model to Assess Oncogenicity of DNA

#### Mouse Strains

- NIH Swiss
   prototype strain used by Burns et al. (1991) for ras studies;
   parent strain of athymic nude strain
- Athymic nude (nu/nu)
   T-cell deficient strain
- K6-ODC constitutively expresses ODC under the keratin 6 promoter; chronic state of tumor "promotional" stimulation
- C57BL/6
   parent strain of K6-ODC mice

#### Construction of pMSV-T24-H-ras and pMSV-c-myc



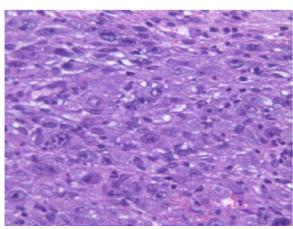
# Tumor Incidence in Mice Injected with ras and myc Expression Plasmids

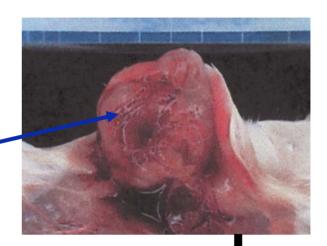
Cohort	ras Plasmid (µg/mouse)	myc Plasmid (µg/mouse)	Swiss		C57BL/6	
			Adult	Newborn	Adult	Newborn
R6	12.5	0	0/10	0/10	0/10	0/8
R5	1.25	0	0/10	0/11	0/10	0/10
R4	0.125	0	0/10	0/9	0/10	0/10
R3	0.0125	0	0/10	0/8	0/10	0/10
RM6	12.5	12.5	2/10	9/11	0/10	2/11
RM5	1.25	1.25	0/10	0/10	0/10	0/10
RM4	0.125	0.125	0/10	0/10	0/10	0/10
RM3	0.0125	0.0125	0/10	0/10	0/10	0/10

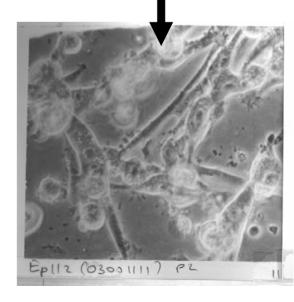
Sub-cutaneous sarcomas developed at injection site at 4 - 7 weeks

#### ras-myc Tumor in NIH Swiss Mouse

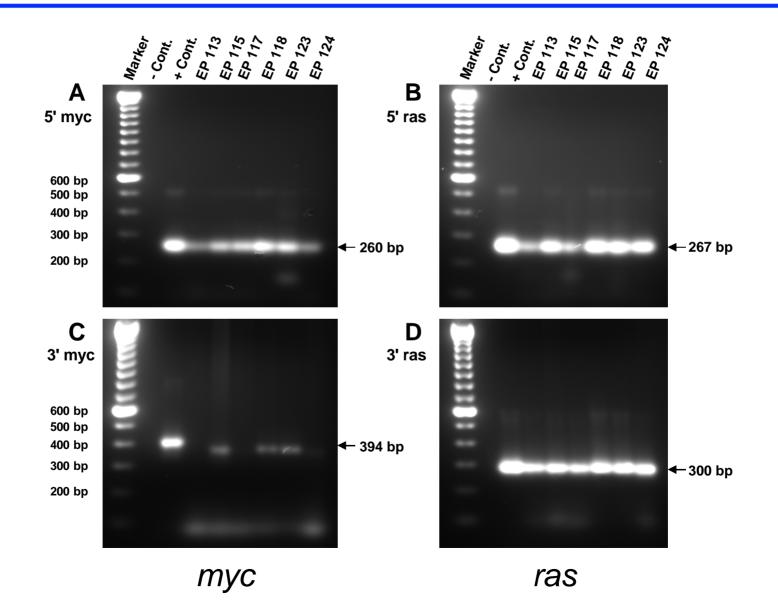








### PCR Analysis of *ras* and *myc* Genes in Tumor Cell Lines



# Summary of the Results: Oncogenicity of DNA

- An MSV LTR expression plasmid has been constructed and tested in vitro for activity (luciferase)
- Expression plasmids for T24 H-ras and murine cmyc under the MSV promoter have been generated and shown to be active in in vitro transformation assays
- The ras and myc plasmids are able to induce tumors in normal mice: NIH Swiss > C57BL/6; newborn > adult
- Cell lines from tumors have ras and myc DNA

#### An in vitro Assay to Assess DNA Infectivity

### Li Sheng and Keith Peden CBER/FDA

Cloned HIV plasmid transfected into 293T cells

Co-culture with Jurkat cells

Follow infectivity with reverse transcriptase activity in medium

#### Calculations of Risk Based on DNA Infectivity - 1 Extrapolations from Viral-Infected Cellular DNA

DNA from HIV-infected cells was tested for infectivity

2.5 μg of this DNA was infectious (in 2 out of 4 experiments; 5 μg in 4 out of 4 experiments)

Assume that between 1 and 2.5 µg infected cellular DNA is infectious

Therefore, in the <u>absence</u> of any treatment (chemical, enzymatic, radiation), residual cell-substrate DNA at **10 ng** could have a safety margin of **100 - 250 fold** 

#### Calculations of Risk Based on DNA Infectivity - 2 Extrapolations from Infectivity of Cloned HIV DNA

Cloned HIV DNA is infectious at 1 pg

HIV proviral genome represents 1.67 x 10<sup>-6</sup> of the diploid cell genome

Therefore, the amount of cellular DNA that would correspond to an infectious HIV dose is:

1 pg ÷ 1.67 x 10<sup>-6</sup> 0.6 μg (600 ng)

Thus, in the absence of treatment (chemical, enzymatic, radiation), **10 ng** of cellular DNA provides a **60-fold** safety factor

## Calculations of Risk Based on DNA Infectivity - 3 Digestion with Omnicleave Nuclease

Digestion of DNA to a mean size of 650 bp resulted in the loss of biological activity of 0.15 µg of pure viral DNA

Based on the proportion of a retroviral genome in the cell, 150 ng of viral DNA corresponds to:

150 ÷ 1.67 x 10<sup>-6</sup> of cellular DNA

 $= 90 \times 10^6 \text{ ng}$ 

= 90 mg

Therefore, for 10 ng of cellular DNA with a single provirus, the safety factor is:

 $9 \times 10^{6}$ 

## Calculations of Risk Based on DNA Infectivity - 4 Inactivation with β-Propiolactone

Treatment of cloned HIV DNA with 0.25%  $\beta$ -propiolactone (BPL) for 72 h at 4°C eliminated the activity of 50 ng

Based on the proportion of a retroviral genome in the cell, 50 ng of viral DNA corresponds to:

 $50 \div 1.67 \times 10^{-6}$  of cellular DNA

 $= 30 \times 10^6 \text{ ng}$ 

= 30 mg

Therefore, for 10 ng of cellular DNA with a single provirus, the safety factor is:

 $3 \times 10^6$ 

## Conclusions from Calculations of Risk Based on DNA Infectivity Studies

- DNA infectivity with HIV-infected cellular DNA and cloned HIV DNA gave similar safety factors for 10 ng DNA 100 - 250 fold with cell DNA, 60 fold with cloned DNA Therefore, the infectivity of a retroviral genome is similar whether integrated or not
- With 10 ng of cellular DNA, nuclease digestion to a mean size of 650 bp could provide a safety margin of 9 x 10<sup>6</sup>
- With 10 ng of cellular DNA, treatment with BPL could provide a safety margin of 3 x 10<sup>6</sup>

These studies suggest ways of obtaining a  $>10^7$  fold margin of safety with respect to residual cell-substrate DNA. *i.e.*, reducing the size of the DNA or inactivating the DNA and having <10 ng DNA per dose.